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(54) Title: METHOD AND MEANS FOR THE PRODUCTION OF HYALURONIC ACID (57) Abstract <p>Supercapsulated strains of group A or C streptococci and the use thereof for the production of hyaluronic acid with a molecular weight exceeding 6 million.</p>		

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Method and means for the production of hyaluronic acid.

The present invention is related to a method for the production of high molecular weight hyaluronic acid by fermentation using supercapsulated strains of streptococci. The invention also relates to a method for the selection of supercapsulated mutants and to mutants producing such hyaluronic acid in high yield.

Hyaluronic acid (HA) or hyaluronan is a glycosaminoglycan consisting of repeating disaccharides of alternating D-glucuronic acid and N-acetylglucosamine molecules. These molecules are joined by a β (1,3)-D linkage while the glucosamine to glucuronic acid linkage is β (1,4)-D.

There are several sources of hyaluronic acid and its molecular weight varies considerably depending on the source. The HA found in synovial fluid has a molecular weight of about 1 to 8 million, that in human umbilical cord has a molecular weight around 3.6-4.5 million and HA in rooster combs may reach very high values, for instance up to 12-14 million, or even higher. The chemical composition of hyaluronic acid is the same regardless of its source and since it is non-immunogenic it has found several applications in medicine (Brimacombe and Webber (1964)). The effectiveness of HA is a result of an unique combination of elastic and viscous properties, which are correlated to the molecular weight. Therefore, there was early an interest in obtaining as high molecular weights as possible.

Accordingly, the literature contains numerous examples of very high values of the molecular weight of HA but these values very often refer to the source material. It should be noticed, however, that since the HA as produced in biological systems like rooster combs, is associated with proteins and other glycosaminoglycans, for example chondroitin sulphate, it has to be extensively purified. Even if very sophisticated methods for purification and sterilisation have been developed it is inevitable that the molecular weight decreases during these steps and the final product in most cases has much lower molecular weight.

The major HA product on the market today is Healon® (Pharmacia AB, Uppsala, Sweden) which has a molecular weight around 3.5 million. This product is prepared from rooster combs according to a method based on the disclosure of US 4141973. From the same source is prepared a HA product with a molecular weight around 5 million, Healon® GV (Pharmacia AB). These molecular weights refer to the sterilized products and this means that the product before the sterilization step must have molecular weights around 5 and 7 million, respectively.

There are very few high molecular weight HA products on the market, in spite of the well-documented usefulness of HA in several medical indications, for instance in ophthalmology. One reason for this is probably the complex purification procedures required in order to obtain a pure product from the sources mentioned above, especially rooster combs, without too much degradation of the molecular chains. Therefore, there is a need for alternative sources or production systems which are well controlled and which allow a simplified purification procedure.

Numerous articles and patent applications have been published which relate to the production of HA in various bacterial systems. The use of bacteria for biotechnological production of HA has been advocated for several reasons, technical, economical as well as ethical. The production by Streptococcus spp. has been known for more than 50 years and most of the systems disclosed seem to refer to group A and C streptococci, for instance encapsulated strains of Streptococcus pyogenes (group A), which is a human pathogen (Kendall et al (1937)), and Streptococcus equi and Streptococcus equisimilis (group C), which are animal pathogens. The synthesis of hyaluronic acid as the major capsular polysaccharide in these pathogens is a way to evade host defenses (Roberts et al (1989)).

The biochemistry of HA synthesis in bacteria involves the action of two, so far known, genes, has A coding for synthase, which is an integral membrane protein and has B coding for UDP-glucose dehydrogenase, which converts UDP glucose to UDP-glucuronic acid. In addition, UDP-glucose needs to be converted to UDP-N-acetyl glucoseamine, which is required for cell wall

biosynthesis (see Dougherty and van de Rijn (1992, 1993) and de Angelis et al (1993)). The control of the synthesis is much less known, for instance what initiates and terminates HA synthesis. However, the stoichiometry of the synthesis provides some guidelines for composition of feed and medium.

The efforts with regard to the development of bacteria-based HA production systems have been focused on the selection of bacteria and suitable culture media. It was early evident that capsulated wildtype strains did not release HA of a molecular weight higher than about 5 million into the fermentation broth, though there were indications in the literature that the actual molecular weight in the capsule might be somewhat higher, see van de Rijn (1983). However, as judged from the literature, including patents and commercially available samples, the molecular weight of bacteria-produced HA is far below that at present produced from rooster combs (see above). It should further be noticed that there often is a very clear difference between high molecular weight values indicated in the literature, which express a desired result, and the values actually obtained.

The highest values obtained in bacteria systems seems to be around 4 million, see for instance
US 4784990 (Bio-Technology General) - HA of 2-3.5 million.
WO9208799 (Fermentech) - HA of 1-3 million,
JP2058502 (Chisso Corp) - HA of 2-3 million,
JP63129991 and JP63028398 (Denki Kagaku Kogyo KK) - HA of 2-4 million, and
EP144019 (Miles Laboratories, Mobay Chemical Corp) - HA of 2-4 million.

It should further be noticed that the values given above refer to HA products which have not been sterilized and it's therefore clear that these materials can not be used for the manufacture of HA products which after sterilization have molecular weights comparable to the Healon® products mentioned above.

All strains of streptococci are aerotolerant anaerobes, that is they are able to grow in the presence of oxygen but they don't use oxygen as electron acceptor. Accordingly, the

discussion or speculation in prior art articles and patents regarding the importance of air doesn't seem to address any parameter of crucial importance for HA production.

Suitable media and conditions for production of HA are discussed in most of the papers related to production and as additional examples of patents or patent applications in this field can be mentioned JP 63141594 and JP 63123392 (Denki Kagaku Kogyo KK) as well as US 4897349 (MedChem Products Inc).

In spite of the numerous publications indicated above there is still a need for an efficient bacteria based production system for high molecular weight HA products. With "high molecular weight" in this connection is meant values exceeding 6 million, in particular over 8 million and especially over 9 million or higher since such a material would be adequate for the manufacture of Healon® GV type products.

I have now found that high molecular weight HA is produced by supercapsulated mutants of streptococcus and one aspect of the invention is the use of such strains in a fermentation system with subsequent purification to obtain HA with molecular weight greater than 6 million, especially greater than 8 or 9 million.

Another aspect of the invention is the preparation and selection of suitable supercapsulated bacteria strains.

The experimental work has been based mainly on the wildtype S. equi ss equi CCUG 22971, which formed mucoid colonies on agar plates and produced HA in liquid medium. From this species acapsular control mutants as well as supercapsulated mutants were derived. Acapsular mutants banded at a density of 1.09 g/cm³, mucoid wildtypes at 1.05 g/cm³ and supercapsulated strains at a density below 1.03, and more precisely about 1.03-1.02 g/cm³ in Percoll gradients (see the experimental part of the description).

The bacterial strains to be used according to the present invention are streptococci, especially of group A and C, and more particularly Streptococcus equi ss equi mutants, which are supercapsulated species having a capsule about at least twice the size of the capsulated wildtype strains as judged from phase-contrast microscopy and india ink staining of cells

growing under optimal conditions, the species banding at a density of or below 1.03 g/cm^3 , for instance in the range of $1.02\text{-}1.03 \text{ g/cm}^3$, and producing HA of a molecular weight exceeding 6 million, especially over 8 or most preferably over 9 million.

The method of producing the bacteria strains comprises the steps of subjecting a bacteria strain, such as a wildtype strain of a Lancefield's group C Streptococci, for instance the at present most preferred strain S. equi ss equi CCUG 22971 to mutagenesis, especially chemical mutagenesis on solid medium avoiding the more cumbersome procedure of mutagenesis in liquid medium, favouring the outgrowth of super mucoid colonies in that the capsule also protects against the mutagenic and toxic chemical and finally enrichment and selection in a density gradient by way of the supercapsulated cells lower density in such a gradient.

Streptococcus equi is a horse pathogen currently grouped together with some other pyogenic and hemolytic Streptococci, which belongs to Lancefield's groups C. Other group C streptococci pathogenic for man or animals have been classified as S. equisimilis, or as S. zooepidemicus mainly from their carbohydrate fermentation pattern. The taxonomic relationships among these strains have not been satisfactorily explored so far and they are only grouped as a taxon Streptococcus sp. (group C) in the First Edition of Bergey's 'Manual of Systematic Bacteriology'. S. dysgalactiae, in contrast, is α -hemolytic and has been recognised as a valid species. It might be most related to S. equisimilis. It has also been proposed that S. zooepidemicus is a subspecies of S. equi. Accordingly, Streptococcus equi should be referred to as S. equi ss equi.

The method of producing HA comprises the steps of (i) selecting a supercapsulated streptococcal strain with the ability to produce HA with molecular weight exceeding 6 million, especially exceeding 8 or 9 million, (ii) cultivating the strain in a bioreactor in the presence of a suitable medium at a temperature below 35°C , preferably in the range of from 30°C to 35°C , especially $31\text{-}33^\circ\text{C}$ and at a pH-value around or below 6.2, such as 5.6 to 6.2 and preferably in the range of

from 5.80 to 5.95, and (iii) purifying the product from the crude mixture.

The medium employed must permit continuous synthesis of hyaluronic acid and not select for non-capsulated cells, which occurs if one tries to optimise the growth rate of cells. It shouldn't contain or release from the reactor any metal ion promoting the degradation of HA, such as iron and copper ions.

The composition of the medium should, in general terms, meet the two requirements of (i) supplying the basic elements (as C, N, O, H, P and S) and necessary growth factors for the build up of the streptococcal cells in correct proportions as well as supply (ii) the elements and compounds for HA-synthesis in sufficient amounts and correct proportions. The composition of any feed should also meet requirement (ii). The compositions of the growth medium was calculated from the composition of microbial cells and the feed composition from the stoichiometry of the HA-synthesis. The basic liquid medium for fermenter cultivations is given in Table I (see also the experimental section, below).

Table I. Compositions of casaminoacids based medium.

Component	Conc. (g/l)	Standard	Range
Bacto Vitamin Assay		12	
Casamino Acids			
Bacto Yeast Extract		3	±1
K ₂ HPO ₄		3	<+11 ^a
Tryptophan		0.4	
MgCl ₂ .6H ₂ O		0.25	
MnCl ₂ .6H ₂ O		0.05	
NaHCO ₃		2	0-2 [*]
Sugar		Glucose 16	±4 ^a
Additions		as specified	

* Depending on the desired pH value

The reactor shouldn't be equipped with any type of baffle or internal component causing extensive turbulence and

agitation must be provided in a very mild way, for instance by gas lift or by any other type of impeller able to achieve good mixing without generating shear forces. This is of crucial importance in order to obtain very high molecular weights and is contrary to the recommendation given in US 4784990, that is "growing with vigorous agitation a microorganism of the genus *Streptococcus*..."

Various culture alternatives have been tested and found to work, for instance batch, fed batch, semicontinuous fed batch and continuous cultivation.

Media

The standard agar plates used were Blood Agar, BA (prepared from horse blood at the Central Bacteriological Laboratory, LU), Bacto Todd Hewitt Agar, THA (Difco) and TYSA made from 10 g/l Bacto Tryptone (Difco), 1 g/l Bacto Yeast Extract (Difco), 1.6 g/l disodium hydrogenphosphate (Merck, PA), 2 g/l sodiumhydrogencarbonate (Merck, PA) 0.1 g/l magnesiumsulphate (Merck, PA), 20 g/l Bacto Agar (Difco) and 8 g/l sucrose (BDH). Liquid medium for initial tests was Todd Hewitt Broth (Difco) supplemented with Bacto Yeast Extract as above.

Mutagenesis

Chemical mutagenesis with nitrosoguanidine (Sigma) was employed (Cerde Olmedo IE and Hanwalt PC (1968)). The wild type strain was spread on a TYSA plate. A few crystals of nitrosoguanidine was applied in the centre. After incubation a clear zone of inhibition was evident around the nitrosoguanidine crystals. Mucoid colonies growing out in the vicinity of the zone edge were selected and subjected to further testing.

Gradient centrifugation

The organisms were harvested after growth in THB by centrifugation, washed once in 0.15 M sodium chloride and resuspended in sodium chloride. Percoll gradients were preformed with 10 ml 25-50 % Percoll in 0.15 M sodium chloride

at 15000 g_{av} for 30 minutes at 4 °C in a fixed angle rotor. Density Marker Beads (Pharmacia LKB Biotechnology AB, Uppsala, Sweden) were added as internal density value standards. A 50 µl volume of the cell suspension was added to each preformed gradient and was then centrifuged at 5000 to 16000 g_{av} for 20 minutes in a swing out rotor at 4 °C (Percoll: Methodology and applications. Pharmacia Laboratory Separation Division).

Estimation of HA molecular weights.

One method routinely used comprises comparative electrophoresis using HA references of various molecular weight prepared from rooster combs (Pharmacia Ophthalmics). The references were diluted to contain about 1.1-1.2 mg/ml and were stored in a freezer at -20 °C. Gels were cast using 0.7-0.9% agarose. The buffer was phosphate-EDTA (2000 ml, 10X contains: Na₂HPO₄ 57.5 g, NaH₂PO₄ 13.1 g, Na₂ EDTA 3.7 g). References and samples were mixed with bromphenolblue/glycerol and applied to the gel. Samples were allowed to enter the gel from the wells at 20 mA constant current and the gel was then run for about 20 hours at 30 V constant voltage. Finally the gel was stained with a solution of Toluidin blue O (0.4%) for 30 min. It was destained in 3% HAC for 15 min and 3-4 times in 1% HAC for 15 min.

Another method employed was SEC-Lalls (Size Exclusion Chromatography-Low Angle Laser Light Scattering). There was good agreement between the two methods up to about 6 million but at higher values the variation was about 10%.

Supercapsulated strains.

A great number of sequences of the steps specified above have been run in order to select a preferred system for the production of high molecular weight hyaluronic acid and it has been found that the basic step is the selection of the bacteria, which must be supercapsulated. This characteristic can of course be described by using various parameters but we have chosen to use the density value at which the selected strains band as a definition of strains according to the invention. The strains band, as discussed above, at a density

equal to or below 1.03 g/cm^3 , for instance in the range of $1.02\text{--}1.03 \text{ g/cm}^3$. This definition is of course is valid also in case other methods than density gradient centrifugation is used for the selection of strains.

Supercapsulated strains also have a highly mucoid colony morphology. When plated on TYSA containing 8g sucrose/l very large ($\gg 5\text{ mm}$ diameter) slimy colonies grow out. The thickness of the capsule as measured in the phasecontrast microscope is much higher for supercapsulated than for capsulated strains. The diameter of the cells is $1.0 \pm 0.2 \text{ }\mu\text{m}$ but the capsule diameter is $\gg 4 \text{ }\mu\text{m}$. The two supercapsulated strains further discussed below, H22 and its derivative H22 NO, are both non-hemolytic and have have been found to produce HA of molecular weights up to about 7.5 and 9.5 million, respectively.

Supercapsulation according to the present invention can furthermore be determined by multi variat data analysis of near infrared spectra of whole cells. Analysis of samples according to this method is a well known technique, see for instance Jolliffe IT (1986), Massart et al (1990), Box et al (1978), Mark and Workman (1991), Marshall and Verdun (1990) and Kalias and Lang (1994).

The first principal component (PC1) correlates to the degree of encapsulation and a supercapsulated strain has a first principal component that is ≥ 0.4 , preferably > 0.5 and especially > 0.7 compared to the first principal component as determined for a weakly capsulated strain exemplified by CCUG 23255, CCUG27365 and CCUG27366 (here referred to as reference strains). The absolute value of this principal component depends on the type of strain. In a test mutant H22 (below) had a first principal component of $+0.3 \pm 0.1$ and the corresponding value of H22NO was $+0.4 \pm 0.05$. Under the same experimental conditions the reference strains had PC1 values of -0.2 to -0.3 .

Sample preparation comprises growth on blood agar at $37 \text{ }^\circ\text{C}$, dissolution of a few colonies in $1.5 \text{ ml } 0.9\% \text{ NaCl}$ whereafter a 100 microliter cell suspension was spread out to $25 \times 25 \text{ mm}$ on an object glass and was allowed to dry in a laminar

flow bench. The NIR spectra were collected by reflectance mode (1100-2500nm) using an InfraLyzer 500, Bran & Luebbe.

Example 1

The supercapsulated mutant S. equi ss equi strain H22 was cultivated by fed batch in 1000 ml volume of medium in a Braun Melsungen Fermenter equipped with a modified impeller having a large surface area. The cultivation temperature was 33 °C and pH was maintained at 6.0 by addition of sodium carbonate solution. The feed was started in the beginning of the log. phase (at 4h) and continued for three hours. The feed rate corresponded to a dilution rate of $D=0.02 \text{ h}^{-1}$. This feed rate is not optimal for maximal molecular weight as was found in other experiments. The medium composition was the one given in Table I above. The feed contained sucrose 25g/l, glucose 10g/l, mannose 0.1g/l, K_2HPO_4 3g/l and yeast extract 4g/l

Table II. Results from fed batch cultivation of strain H22

Time (h)	OD ₆₂₀	Capsule	Molecular Weights (10^{-6})		Conc HA (mg/l)
			E-fores	SEC-LALLS	
0	0.146	-ND	ND	ND	ND
4	0.409	++	ND	ND	ND
7	0.87	+++	6.8	7.1	226
12	1.13	++	6.8	7.1	376
14	1.15	++	6.8	7.4	416
24	1.11	(+)	6.3	7.5	390
26	1.10	-	6.5	6.4	316

ND=not determined.

Example 2

The strain S. equi ss equi H22 was cultivated in an airlift

reactor at a temperature of 37 °C using the tryptone based medium (concentrations in g/l):

Tryptone	8
Yeast extract	3
NaCl	2
K ₂ HPO ₄	3
MgCl ₂	0.25
MnCl ₂	0.12
NaHCO ₃	2
Glucose	12

When growth had started a "feed" with the composition (conc. in g/l):

Yeast extract (3), Tryptone (8), K₂HPO₄ (5) and Sucrose (350)

was added.

The "feed" volume was 1100 ml which was added during 10 hours. The operating volume of the reactor was 4500 ml and it was kept constant by a pump connected to a level tube and operating with a high speed. The pH-value was kept at 7.1 by addition of 2M Na₂CO₃ during the period of semicontinuous operation. The air was then turned off but the development in the reactor followed for a further 24.5 hours, mainly in order to monitor the degradation of HA. An analysis of the most interesting parameters gave the following results:

molecular weight (MDa)	6.3 (max. value)
degradation rate (MDa/h)	0 (the feed phase)
	0.019 later
viscosity at 12 h	1.504 and
HA content (mg/l)	800 (max value).

The molecular weight was accordingly above 6 million although neither feed rate nor pH was optimal for high molecular weight formation.

Example 3. A continuous cultivation study on the supercapsulated strain H22 NO.

In this experiment a temperature of 33 °C, a pH of 6.0 and a constant dilution rate of 0.10 h⁻¹ was used. The media used in this experiment at the different steady states varied as follows:

S.s. No.	Glucose (g/l)	Phosphate (g/l)	PO ₄ /Glu	Yeast extr. (g/l)	YE/Glu	Add. (mg/l)
1	16	3.2	0.2	2.4	0.15	-
2	16	6.4	0.4	2.4	0.15	-
3	16	6.4	0.4	2.4	0.15	UDP(10)
4	16	8	0.5	2.4	0.15	-
5	20	8	0.4	2.4	0.12	-
6	20	8	0.4	2.4	0.12	RIB(2)

and the results of continuous cultivation were:

S.s. no.	NU	OD	TS (g/l)	M.w. (MDa)	HA-konc. (mg/l)	S _{out} (g/l)
1	240	1.36	14.1	7.7	142	0.29
2	228	1.30	15.1	8.7*	109	0.29
3	208	1.20	15.4	9.1	124	0.26
4	224	1.40	17.4	9.1	145	0.28
5	288	2.45	18.6	7.1	145	0.28
6	292	3.31		7.0	159	0.29

* an extensively purified sample from this steady state gave a m.w of 6.1 by Lalls

It is evident from the results that the molecular weight in the different steady states was high with 9.1 MDa as the max. value. The yield in this specific example was rather low but in another series of experiments values up to about 350 mg/l have been reached. An increase in the level of phosphate doesn't give a higher yield, but an increase in the molecular weight is observed instead, demonstrating the finding that there is often

an inverse relation between yield and molecular weight.

During one week of cultivation the strain was stably non-hemolytic.

It is clear from the experiments indicated above that the new superencapsulated strains are able to produce HA of much higher molecular weight than what has earlier been achieved in bacterial systems. A very promising tool for production of HA has accordingly been developed.

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Claims.

1. Supercapsulated strain of streptococci producing hyaluronic acid with molecular weight exceeding 6 million.
2. Strain according to claim 1 characterized in that it is a group A or C streptococci.
3. Strain according to claim 1 or 2 characterized in that it bands at a density of ≤ 1.03 g/cm³ in a Percoll gradient.
4. Strain according to claim 3, characterized in that it bands at a density in the range of 1.02-1.03 g/cm³ in a Percoll gradient.
5. Method of producing high molecular weight hyaluronic acid comprising the steps of
 - (i) selecting a supercapsulated strain of streptococci having the ability to produce hyaluronic acid with molecular weight exceeding 6 million,
 - (ii) cultivating said strain at a temperature of 30 to 35 °C in a reactor under agitation conditions substantially free from shear forces and in a culture medium having a pH in the range of from 5.6 to 6.2,
 - (iii) purifying the HA formed in step (ii) from the culture medium to give said HA with molecular weight above 6 million.

6. Method according to claim 5, wherein the strain is a group A or group C streptococci

7. Method according to claims 5 or 6 wherein the supercapsulated strain has been obtained by mutagenesis.

8. Hyaluronic acid having a molecular weight of more than 6 million produced by a supercapsulated bacteria strain.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/SE 95/00585

A. CLASSIFICATION OF SUBJECT MATTER

IPC6: C12P 19/26, C12N 1/20

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

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IPC6: C12P, C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

MEDLINE, CA, WPIDS, IFIPAT

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 4517295 A (JAMES W. BRACKE ET AL), 14 May 1985 (14.05.85), see claims and column 4, lines 64 - 68 --	1-2,5-8
X	CA 1328841 A (BROWN, KAREN K. ET AL), 26 April 1994 (26.04.94), see claim 11 --	1-2,5-8
X	US 4782046 A (KAREN K. BROWN ET AL), 1 November 1988 (01.11.88), see claims -- -----	1-2,5-8

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Information on patent family members

International application No.

PCT/SE 95/00585

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US-A- 4517295	14/05/85	AU-B- 556730 AU-A- 2076883 CA-A- 1212645 EP-A,B- 0137786 SE-T3- 0137786 WO-A- 8403302	13/11/86 10/09/84 14/10/86 24/04/85 30/08/84
CA-A- 1328841	26/04/94	NONE	
US-A- 4782046	01/11/88	AU-B- 573768 AU-A- 3580684 CA-A- 1270219 EP-A,A,A 0144019 SE-T3- 0144019 JP-A- 60133894 US-A- 5316926	23/06/88 30/05/85 12/06/90 12/06/85 17/07/85 31/05/94

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INTERNATIONAL SEARCH REPORT

International application No.

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Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 1-2
because they relate to subject matter not required to be searched by this Authority, namely:
Molecular weight is not an acceptable distinguishing feature as it only defines the problem but not the solution. The wording "supercapsulated" has only been shown to solve the problem in certain cases but not that the feature itself is the solution. The consideration on inventive step given is based on the claims, not being clear and concise.
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

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